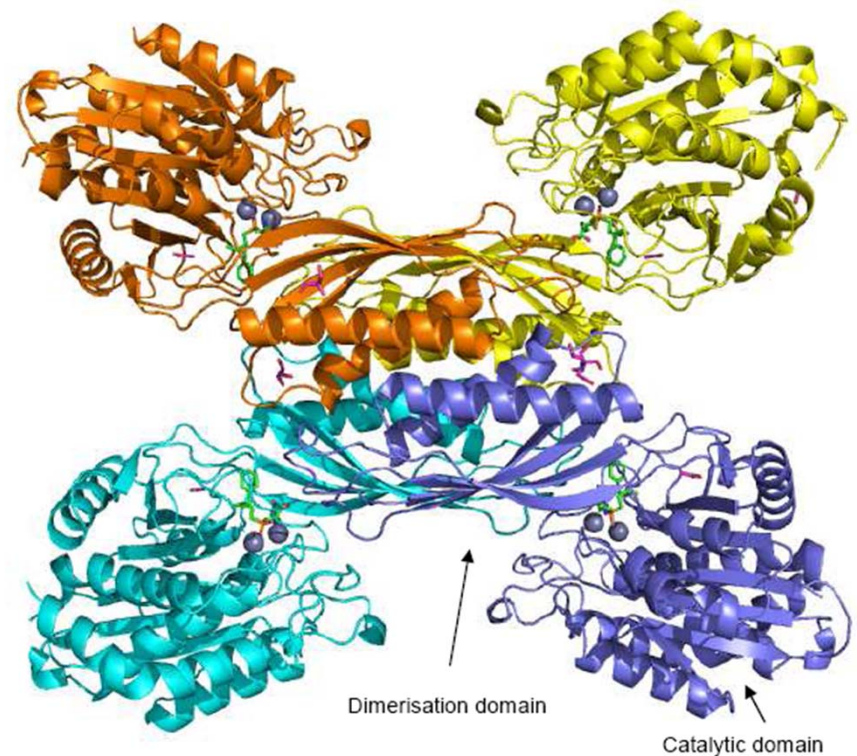


Blocking AMRE – Chemistry and Biochemistry at a Nasty Enzyme

Dr. Andreas Natsch

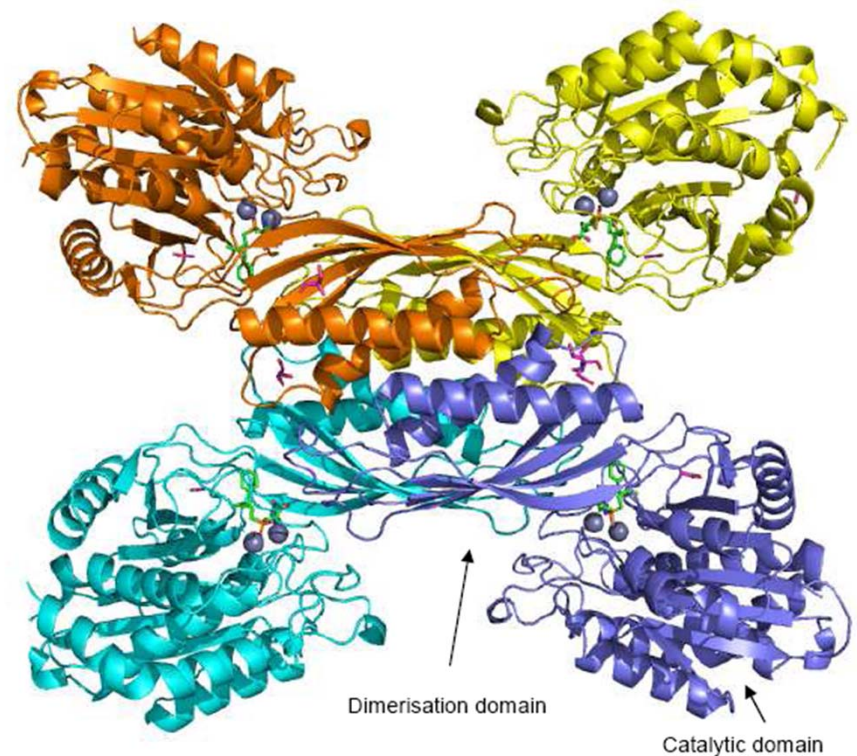
Dr. Fridtjof Schröder

Givaudan Schweiz AG, Dübendorf



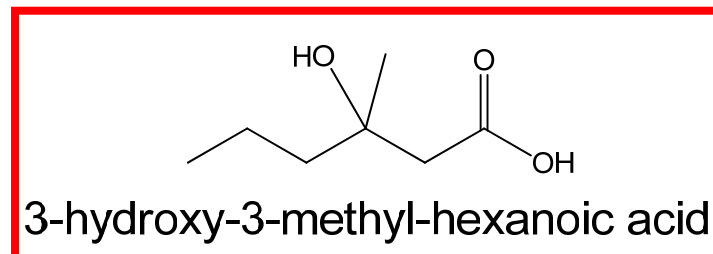
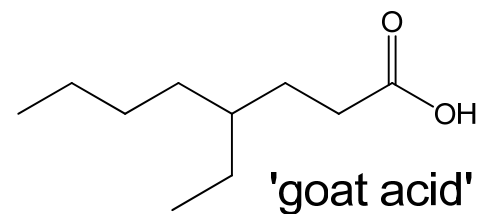
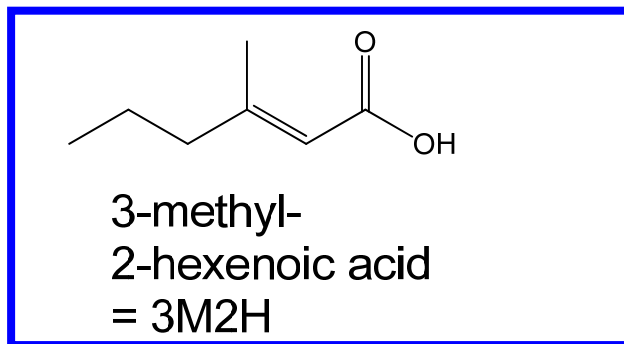
Malodour counteragents: AMRE inhibitors

- **Sweat – composition and biosynthetic origin**
- **AMRE inhibitors: intervention strategy**
- **Synthesis of inhibitors**
- **Inhibition rates and structures**
- **Biochemical and clinical studies**



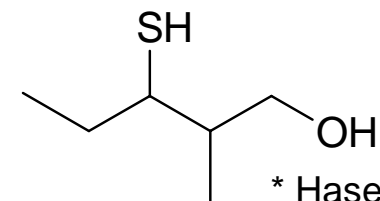
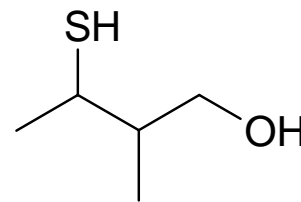
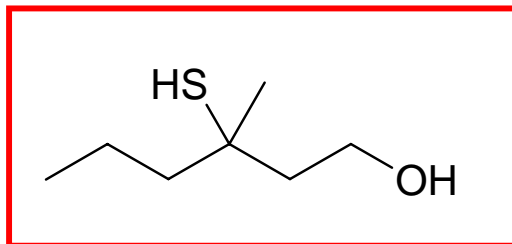
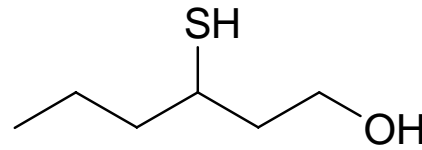
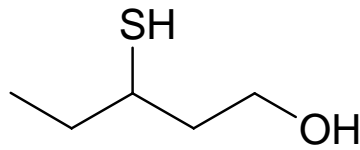
The chemistry of human axilla odors: 1. Volatile carboxylic acids

- 3-methyl-2-hexenoic acid long known as body odorant of schizophrenics
- Later found in the general population
 - ◆ Along with other carboxylic acids such as goat acid
- 3-hydroxy-3-methyl-hexanoic acid quantitatively most abundant body odorant



2. Sulfur volatiles

- Three research groups* reported in 2004 sulfanylalkanols as a further structural class
- 3-sulfanyl-3-methyl-hexanol is the most abundant compound



* Hasegawa *et al.*
 Starckenmann *et al.*
 Natsch *et al.*

The biochemical formation of axilla odors

- First observations, Shelley et al., 1953 stated:

‘No odor could be detected in apocrine sweat after collection’

*‘**Bacterial action** is necessary for the production of odor from apocrine sweat’*

- Leyden et al:

*‘High level of body odor is associated with **large population of Corynebacteria** in the axilla’*

CONCLUSION:

⇒ Sweat contains a non-odoriferous **‘precursor molecule’**

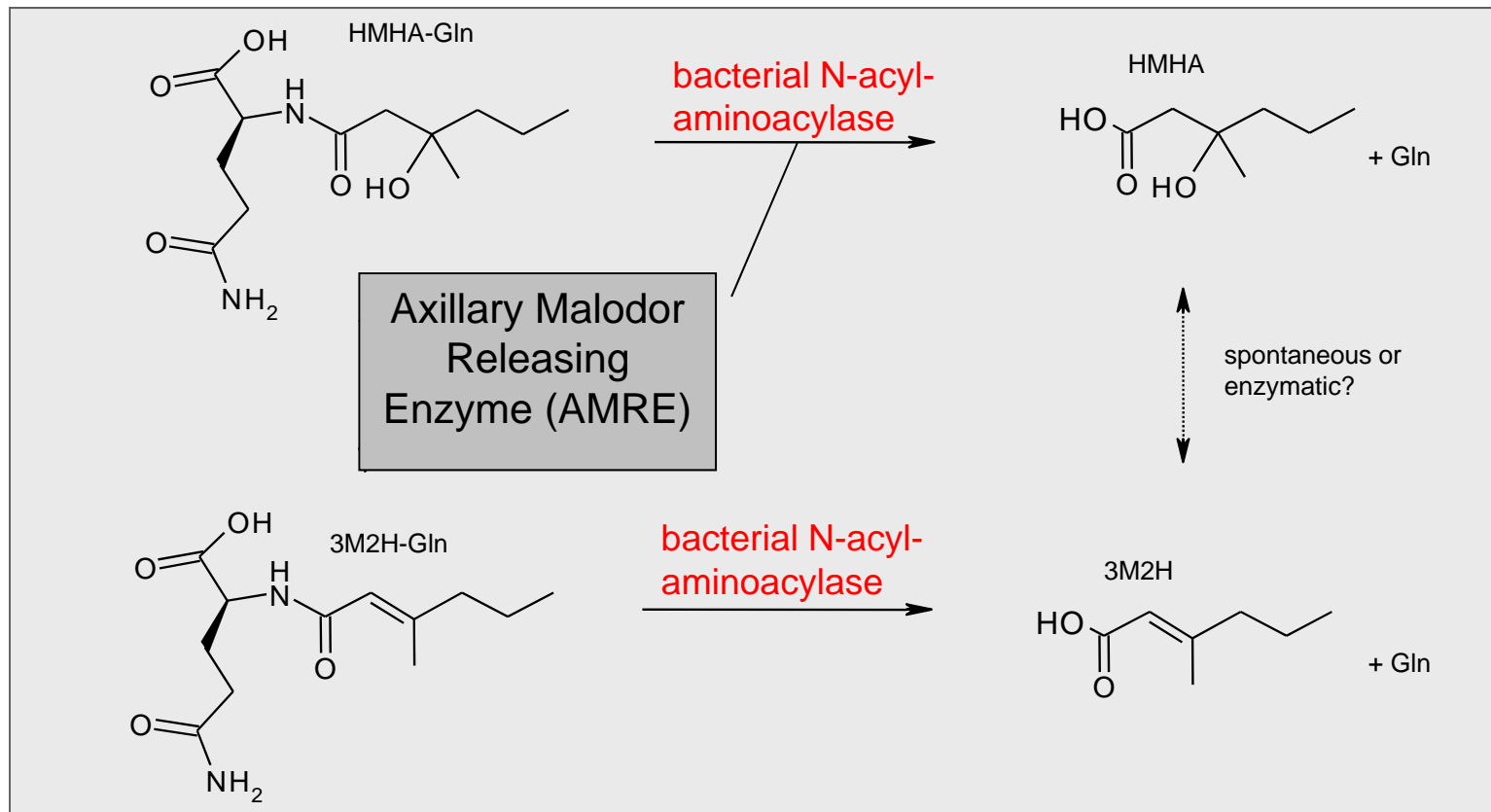
⇒ Skin inhabiting Corynebacteria have **enzymes** which **transform the precursor** into the odorant

Biochemistry: two key questions

- I. Chemical structure of the secreted odor-precursors ?
- II. Which bacterial enzymes cleave the precursors?

Precursors for acids

- The malodor acids in fresh sweat are covalently linked to a glutamine residue



Axillary malodor releasing enzyme in skin bacteria

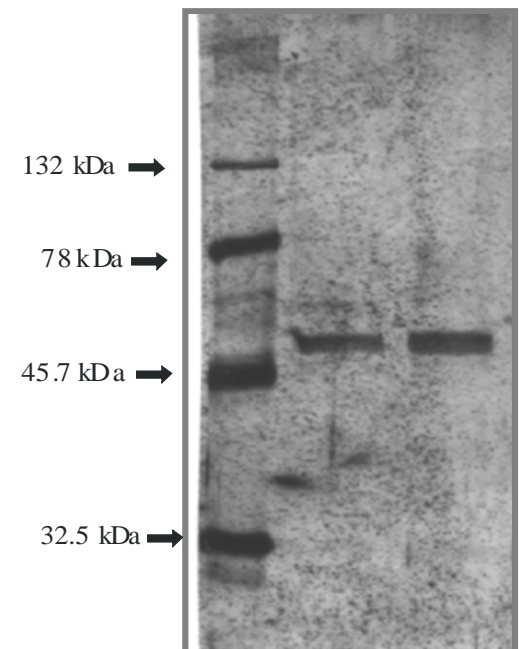
- Cleavage of malodor precursor

strain	species assignment	release of 3M2H from 3M2H-Gln	release of HMHA from HMHA-Gln
Ax7	<i>Corynebacterium group G</i>	<0.005	<0.005
Ax19	<i>Corynebacterium jeikeium</i>	1.132	0.735
Ax20	<i>Corynebacterium striatum</i>	0.217	0.200
Ax21	<i>Corynebacterium bovis</i>	0.449	0.037
Ax1	<i>Staphylococcus capitis</i>	<0.005	<0.005
Ax6	<i>Staphylococcus epidermidis</i>	<0.005	<0.005
Ax9	<i>Micrococcus luteus</i>	<0.005	<0.005

- Malodor is mainly released by *Corynebacteria*
- ⇒ Subjects with high population of *Corynebacteria* have strong body odor

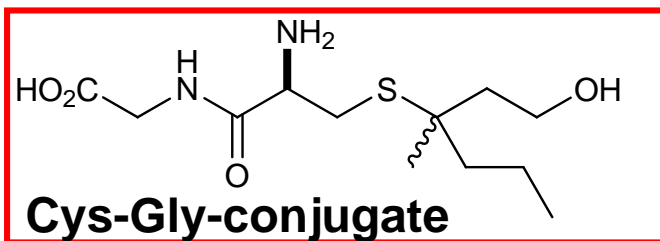
Cloning of the gene coding for AMRE

- Enzyme purified from cellular extracts of *Corynebacterium Ax20* with 4 chromatographic steps
- Partial amino acid sequence determined
- Full length gene cloned by degenerated PCR and chromosomal walking
- Protein expressed in recombinant *E.coli*
- Fluorescent high throughput *in vitro* screening assay developed

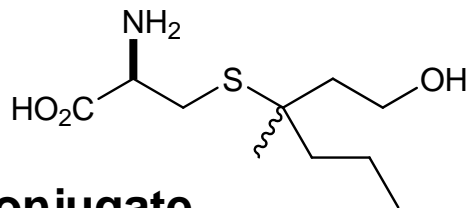


Biochemistry of release of sulfur compounds

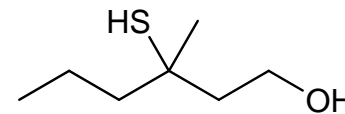
- **Focus of today's talk is AMRE** (Glutamine-aminoacylase), but we also identified two enzymes involved in sulfur release.....
- 3-sulfanyl-3-methylhexanol linked to Cys of Cys-Gly dipeptide*, typical degradation product of a glutathione-adduct



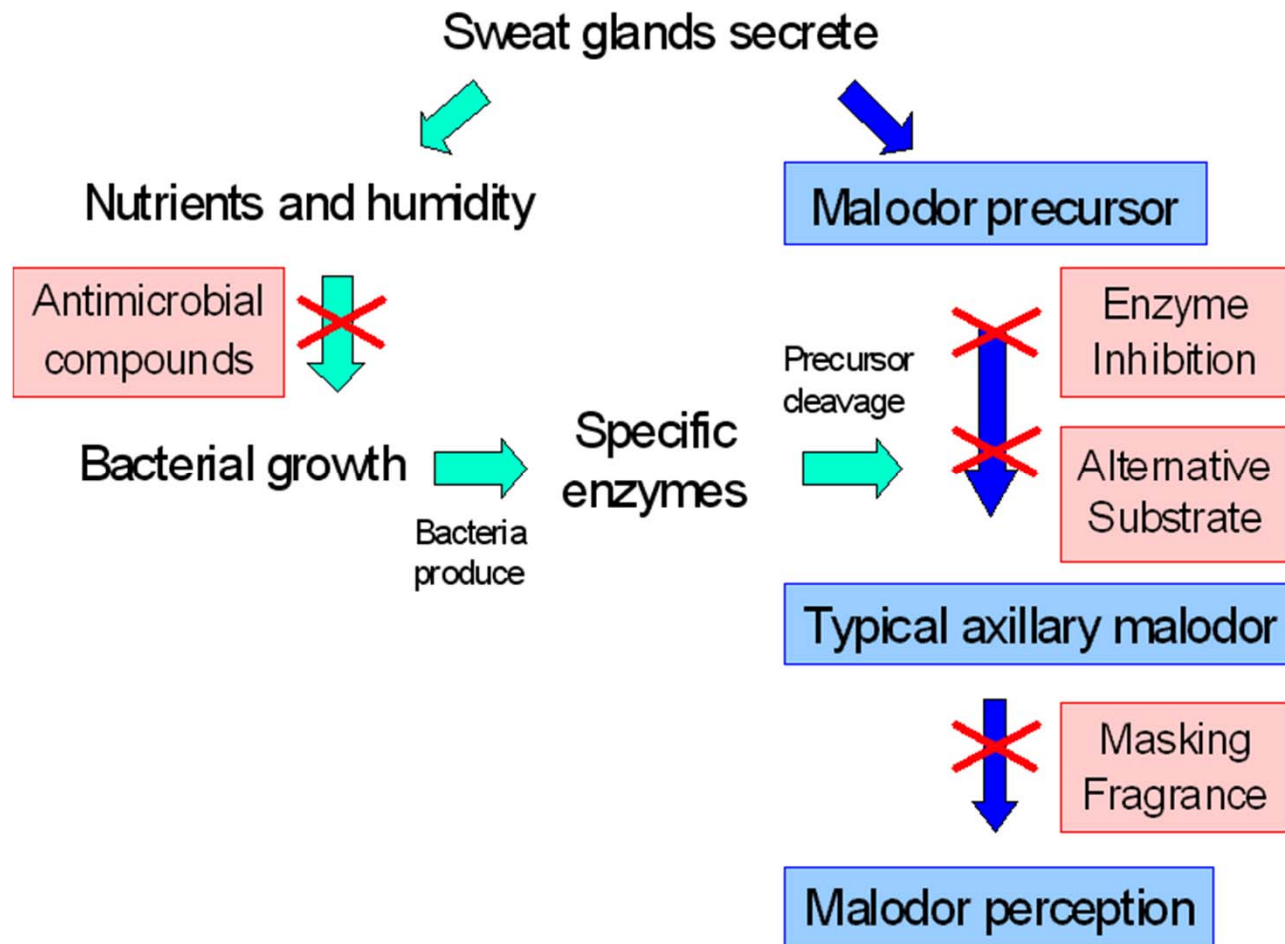
Novel dipeptidase from Ax20



cystathionine-β-lyase from Corynebacterium Ax20

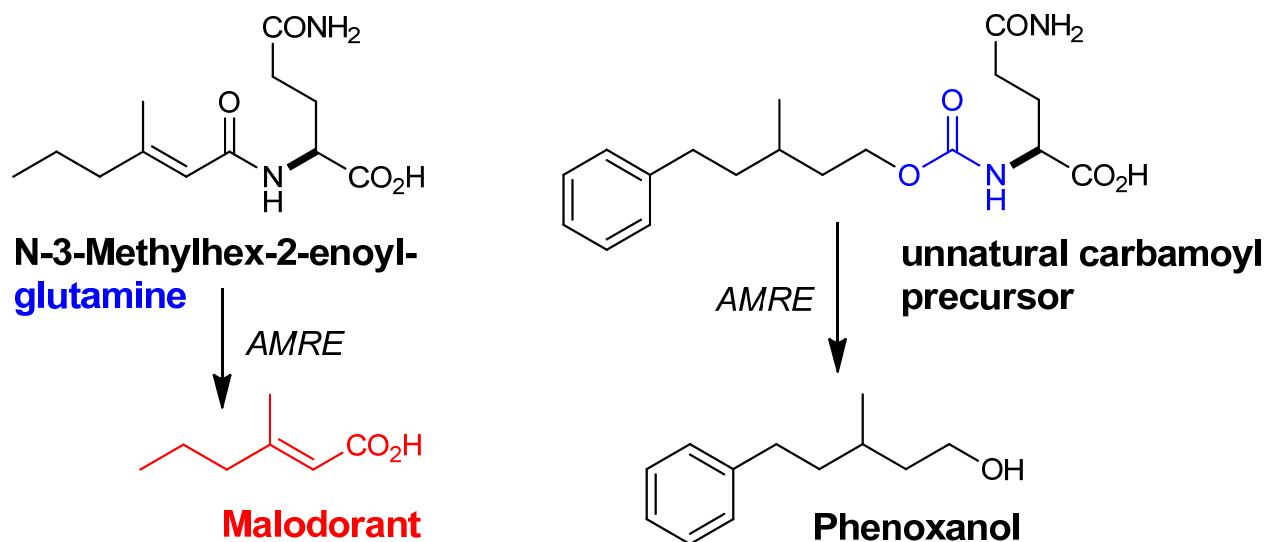


Intervention strategies for deodorant ingredients



AMRE: replacement of the malodorant by a fragrance ingredient

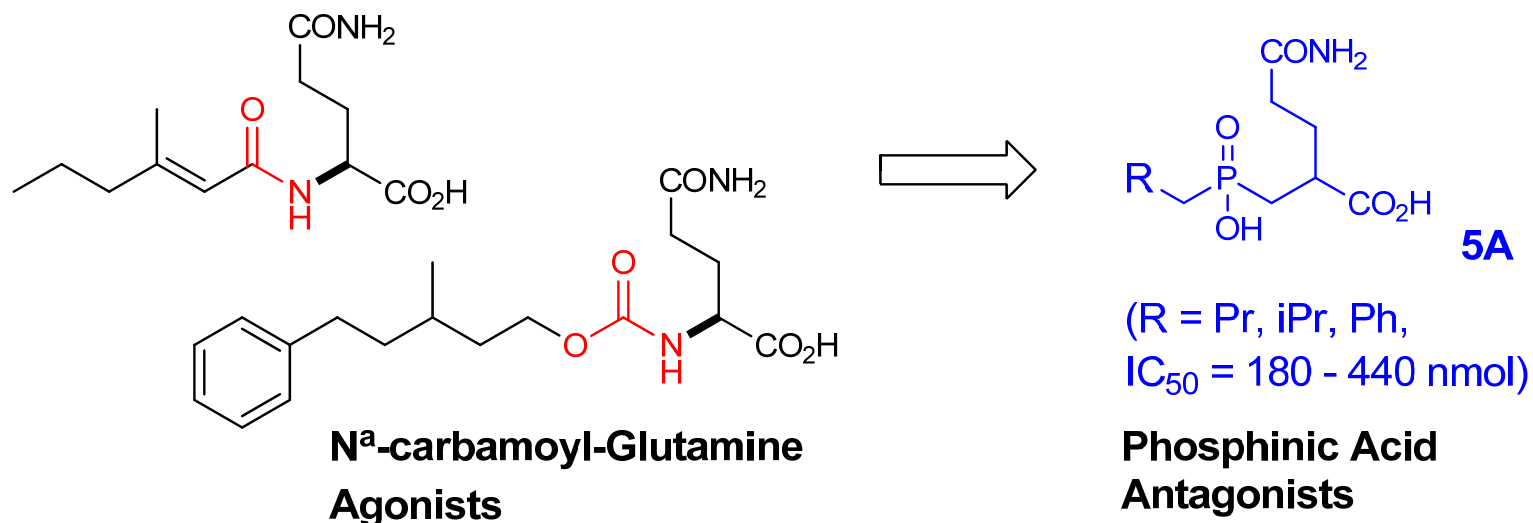
- **Unique substrate specificity: only N^α-substituted Glutamines are recognized, closely related amino acid derivatives not**
- **A large range of different hydrophobic N^α-substituents is tolerated: amides, carbamates...**



- **Disadvantage: an excess over the natural substrate is needed**

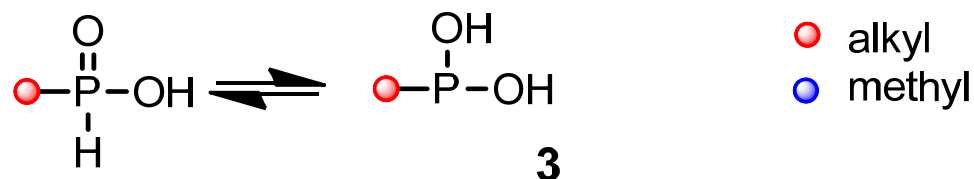
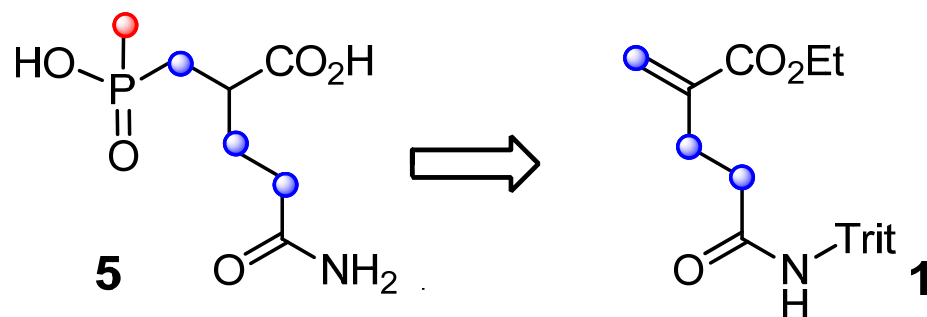
AMRE inhibitors:
replacement of the carbamoyl moiety
by a non-cleavable group

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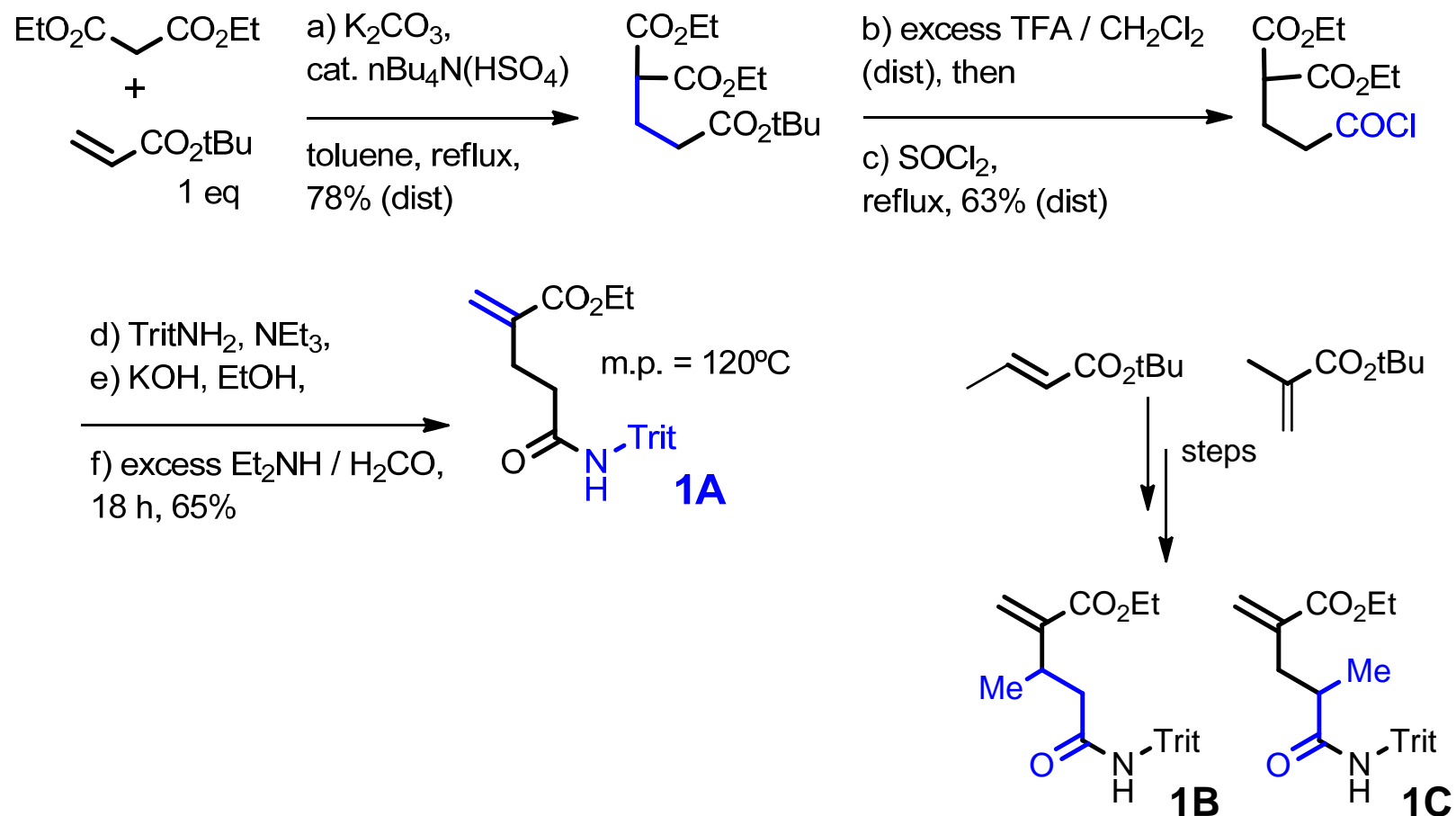
- A first generation of phosphinic acid analogues was synthesized and found to be promising AMRE inhibitors.
- IC₅₀ = concentration of an inhibitor needed to reduce the enzymatic activity by ½ at a given substrate concentration

Phosphinic acid analogues of N^α-substituted glutamines: modifications and building blocks

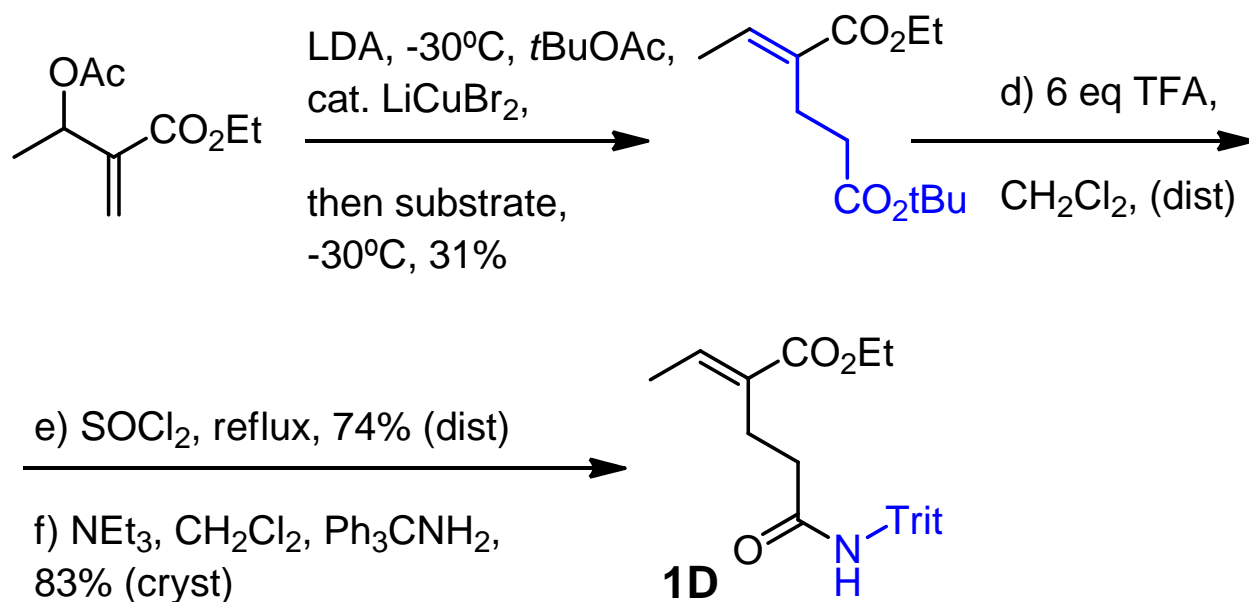


- N-Trityl amide = “solid phase”
- Michael addition of alkylphosphinic acids

Preparation of building blocks 1A-C (0.5 – 1 kg scale)



Preparation of building block 1D

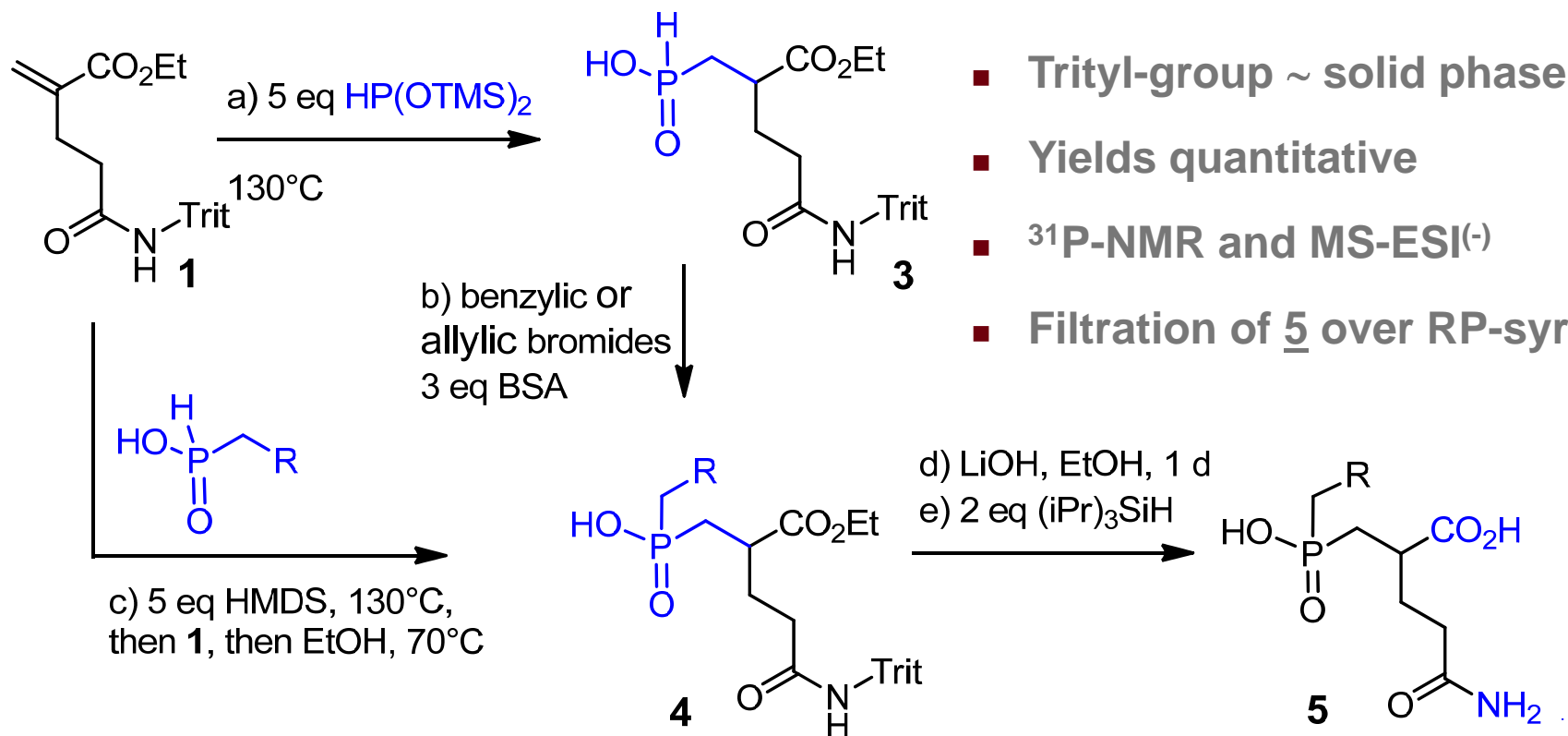


Substrate : C. Yu, B. Liu, L. Hu, *J.Org.Chem.* 2001

Allylic substitution: J. Villieras et al., *J.Organomet.Chem.* 1990

Phosphinoyl-AMRE inhibitors: synthesis

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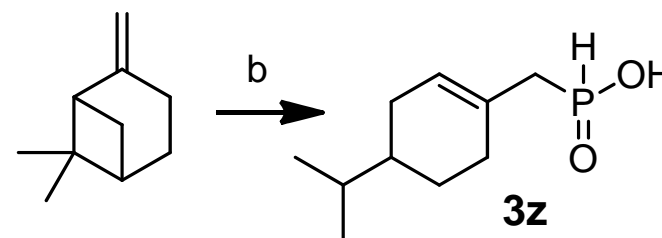
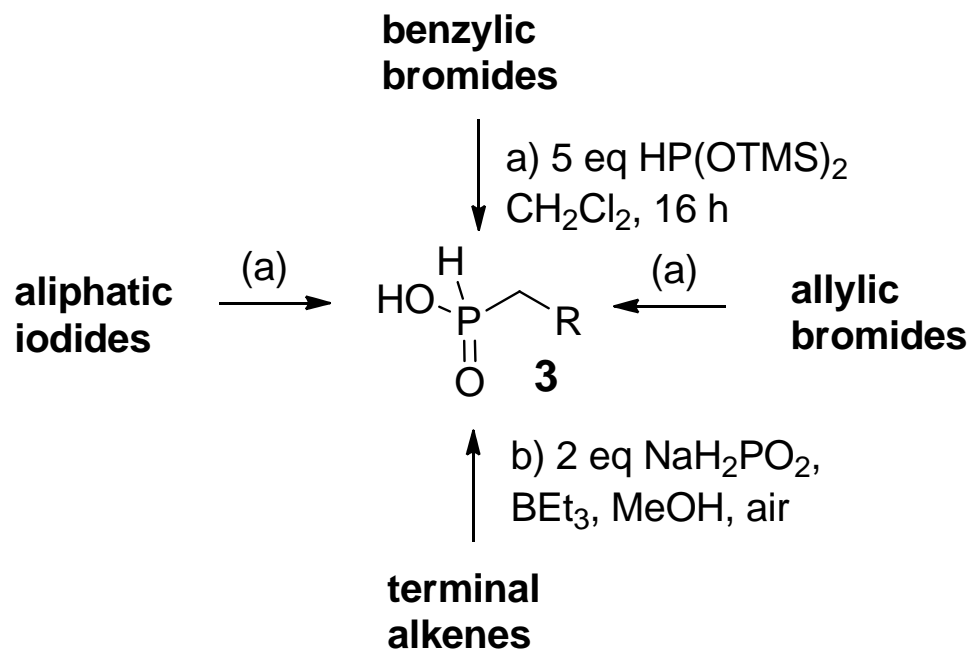
C-P Coupling Steps:

a) Michael addition of $\text{HP}(\text{OTMS})_2$: W.P.Malachowski, J.K.Coward *JOC*, 1994

b) Allylation, benzylation: L.A.Reiter, B.P.Jones, *JOC*, 1997

c) Michael addition of alkyl phosphinic acids: D.Georgiadis, A.Yiotakis, *Tetrahedron*, 1999

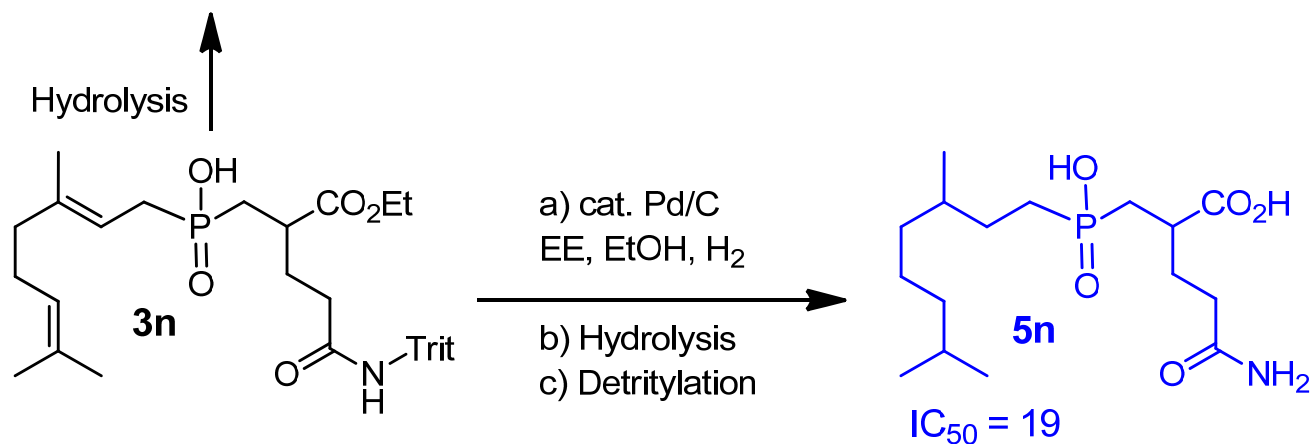
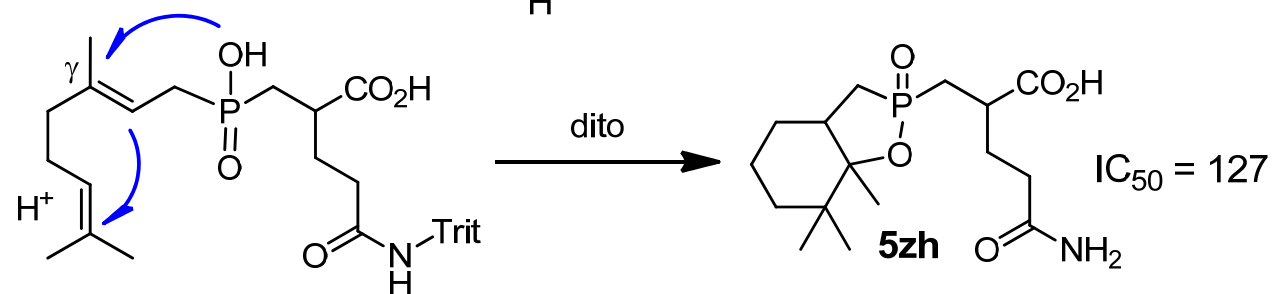
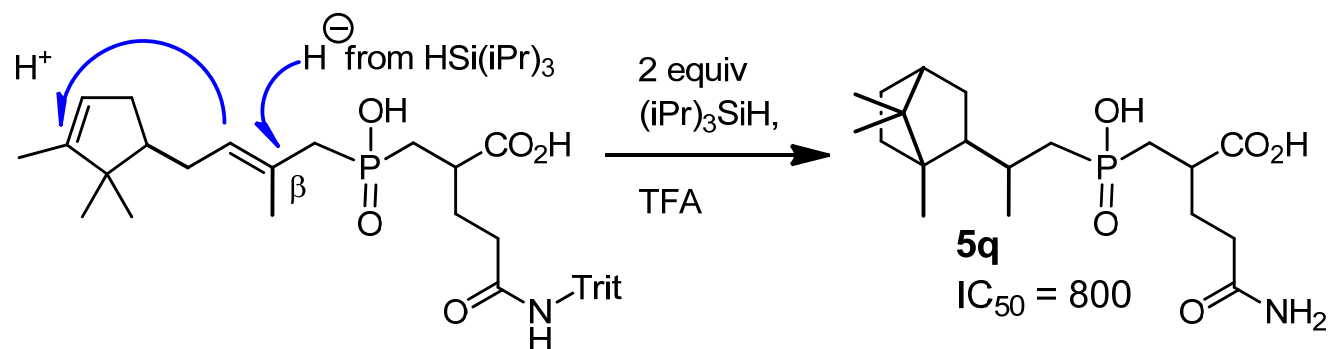
Alkylphosphinic acids: synthesis



a) H.An et al. *JOC* 2001
a,b) S.Deprele, J.-L.Montchamp, *JOC* 2001

Free radical rearrangement:
D.R.Battiste, D.L.Haseldine,
Synth. Commun. 1984

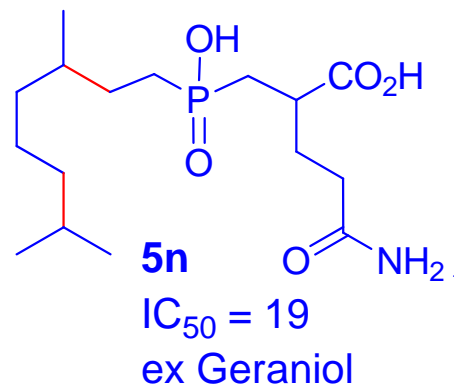
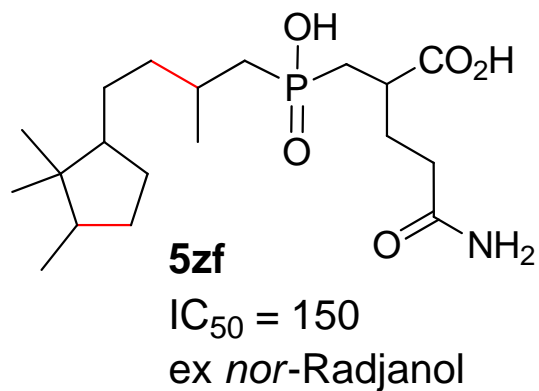
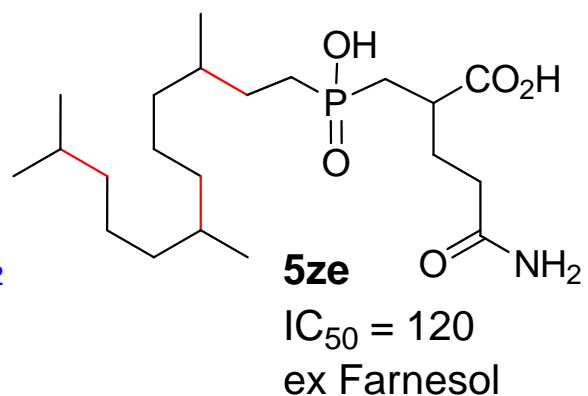
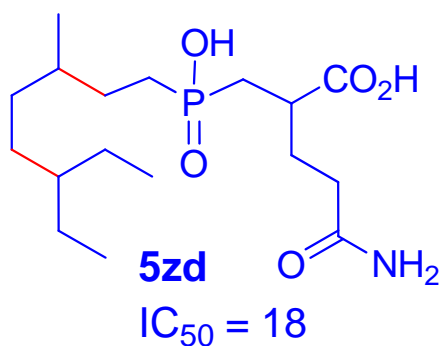
Carbocationic rearrangement during de-tritylation



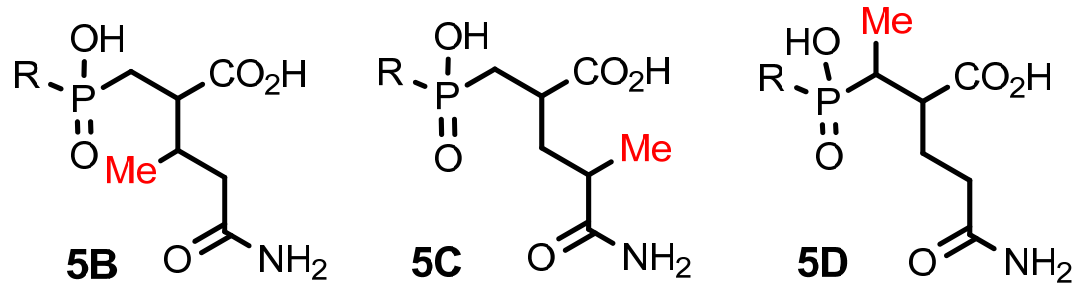
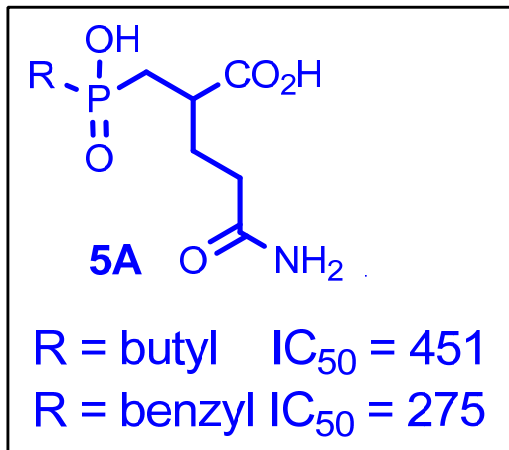
Allylic precursors:

β -substituents fine,
but γ - need prior
hydrogenation of the
 β,γ -double bond

Inhibitors: via hydrogenation
of allylic phosphinyl precursors



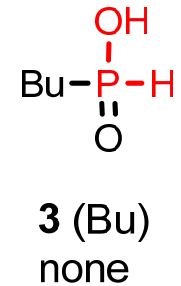
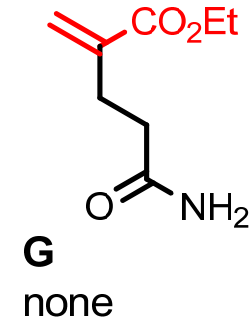
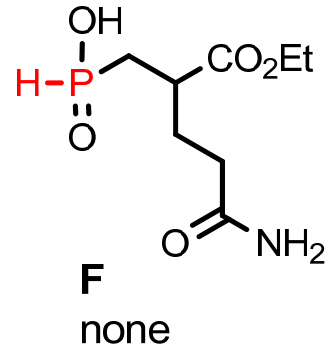
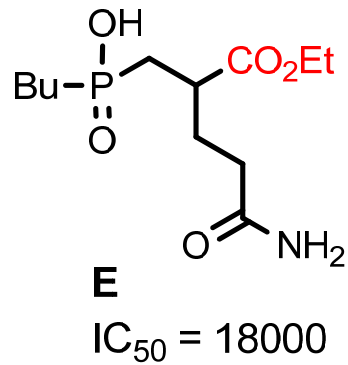
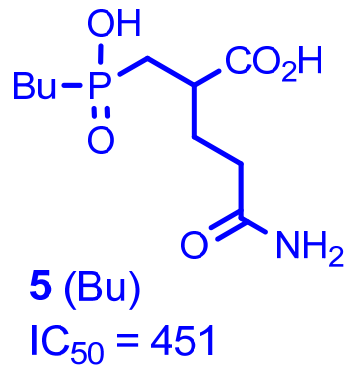
Inhibition: methyl-groups at the backbone
 Comparison with first generation inhibitors



no inhibition

Compounds with methyl-groups at the backbone do not inhibit ($IC_{50} > 40000$)
 in contrast to inhibitors with unsubstituted glutamine backbones

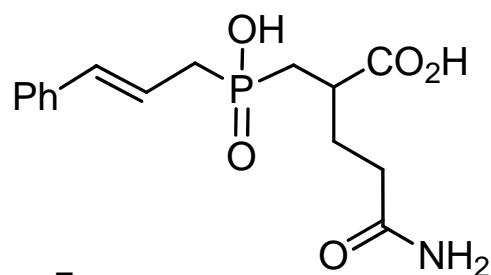
Inhibition: reference compounds



Glutamine-type backbone + dialkyl phosphinyl group necessary
 as well as free CO_2H , free amide

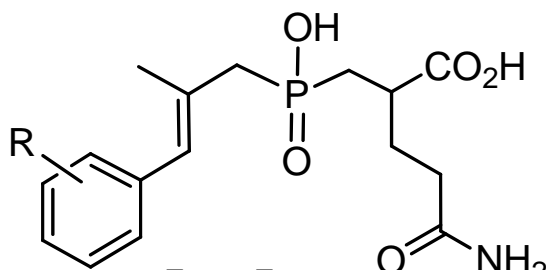
Inhibition: P-allylic analogues:
were good but not better than 1th generation inhibitors,
Exception 5z

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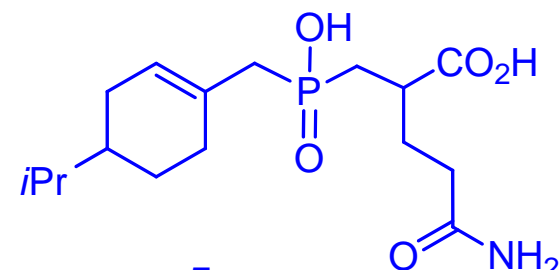
5m

IC₅₀ = 1167



5o - 5s

IC₅₀ = 460 - 2500

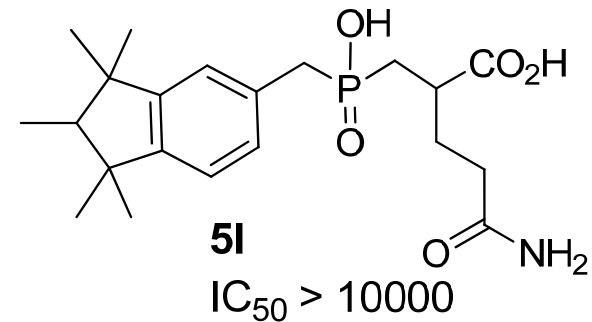
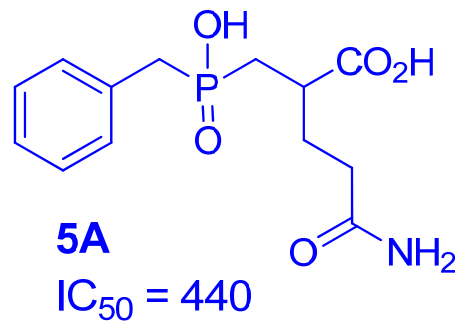
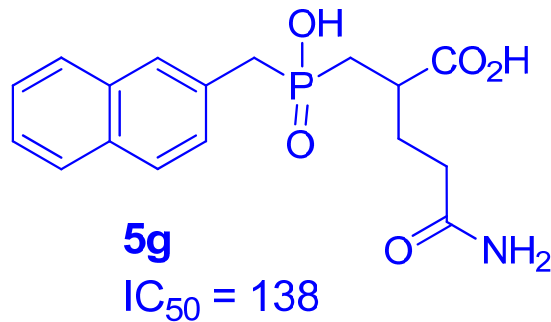


5z

IC₅₀ = 32

9 examples: R = *ortho*- and *para*-tBu, *iPr*, *iBu*

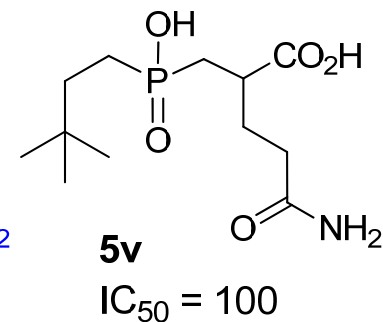
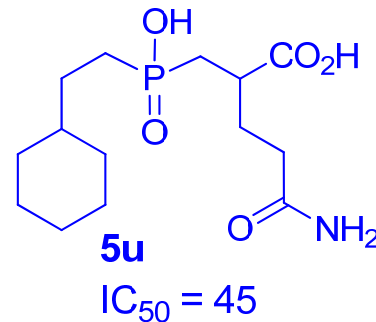
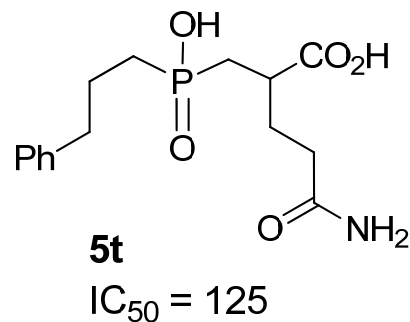
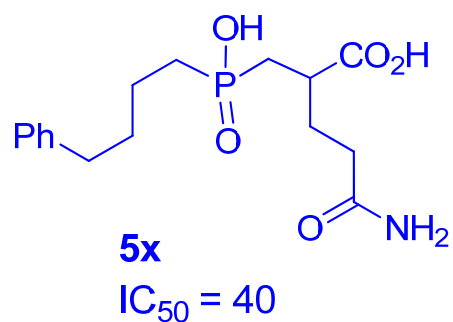
Inhibition: P-benzylic analogues:
 some were slightly better than the 1st generation
 P-Benzyl inhibitor



9 examples: +I and -I substituents (fluorination):

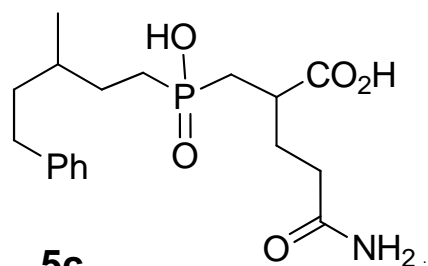
steric rather than electronic effects, planarity

Inhibition: P-Alkyl analogues:
good to very good inhibition

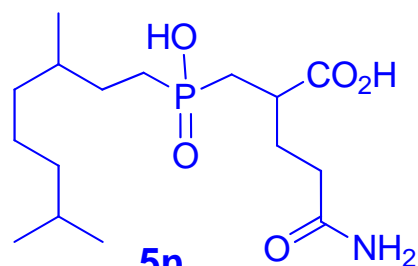


8 examples: flexibility in the α,β -position, P-ethylene bridge

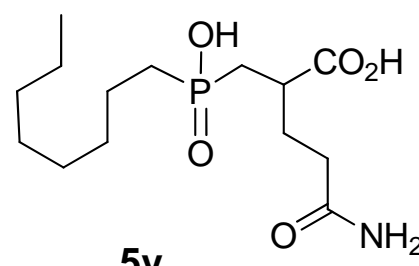
Inhibition: 2th generation



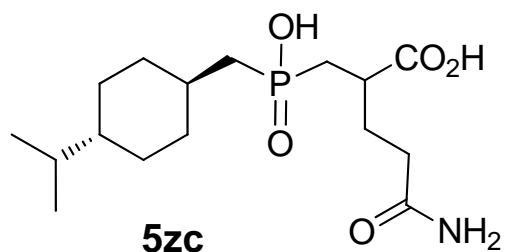
5c
ex Phenoxanol



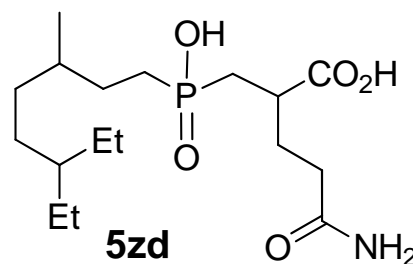
5n
ex Geraniol



5y
ex octyl iodide



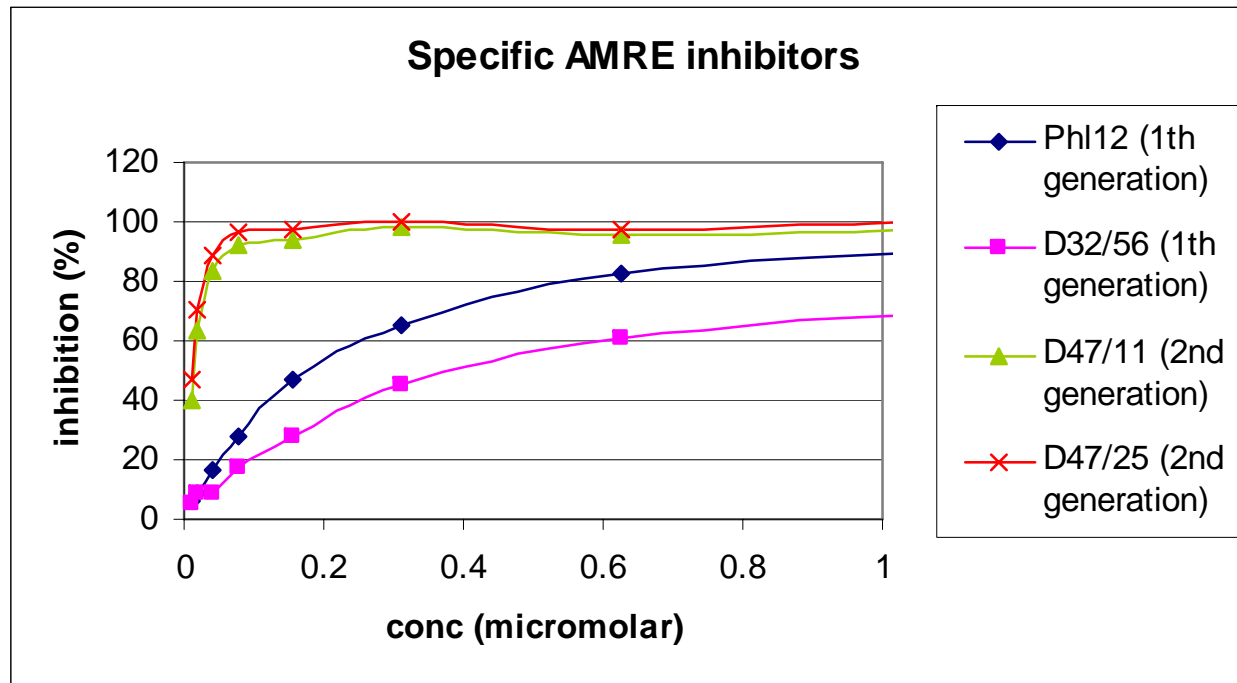
5zc
ex Pinene



5zd

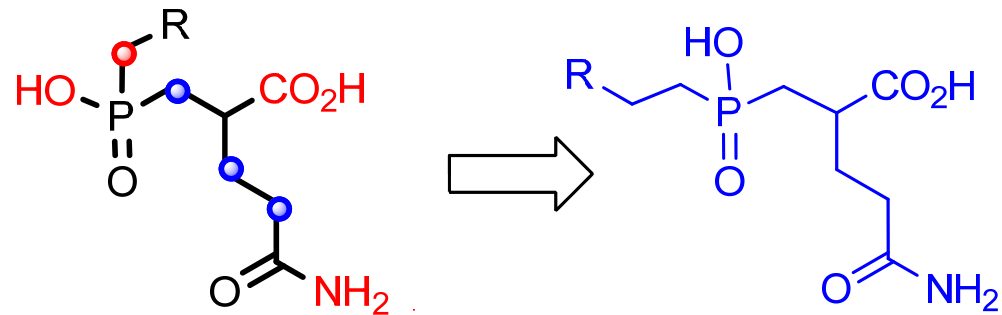
5 best examples: IC₅₀ = 11 - 19

Inhibition: 2th generation



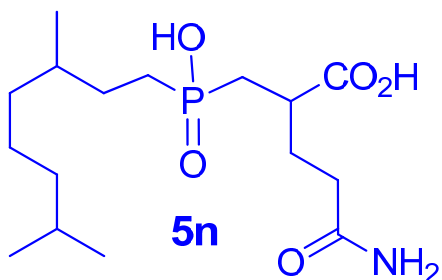
- Second generation: $IC_{50} \sim 10 - 20$ (versus $200 - 400$, 1th generation)

Inhibition results:
elaboration of the key



- Glutamine backbone
- Substituents at the backbone detrimental
- Requirements: dialkyl phosphinoyl group, free CO₂H, free amide
- Alkyl phosphoryl side chain
 - P-ethylene bridge (exception β-alkyl)
 - Allylic DB needs prior hydrogenation
 - P-alkyl > P-allyl > P-benzyl

Lead compound 5n



$IC_{50} = 19$

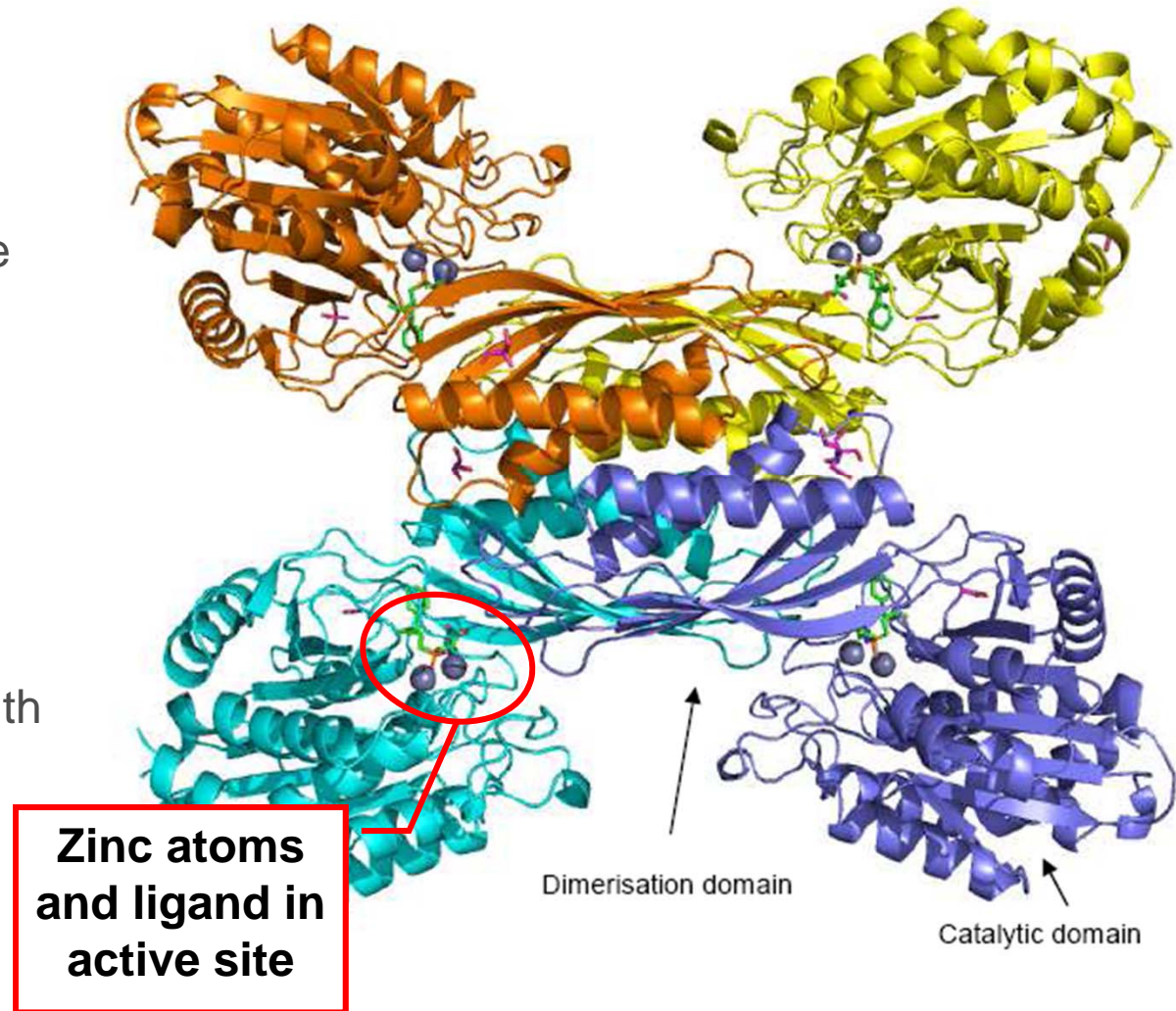
mp 135°-145°C (iPrOH, EE 1:2)

> 98% purity (NMR, ESI-MS)

- 75g prepared in 11 steps starting from 1 kg malonate and acryl ester.
- Optimization: Volume yields, steps combined, RP-filtration of the final product, crystallization.
- Toxicological Testing: AMES (mutagenicity), Acute Toxicity, LLNA
→ no toxicological effects

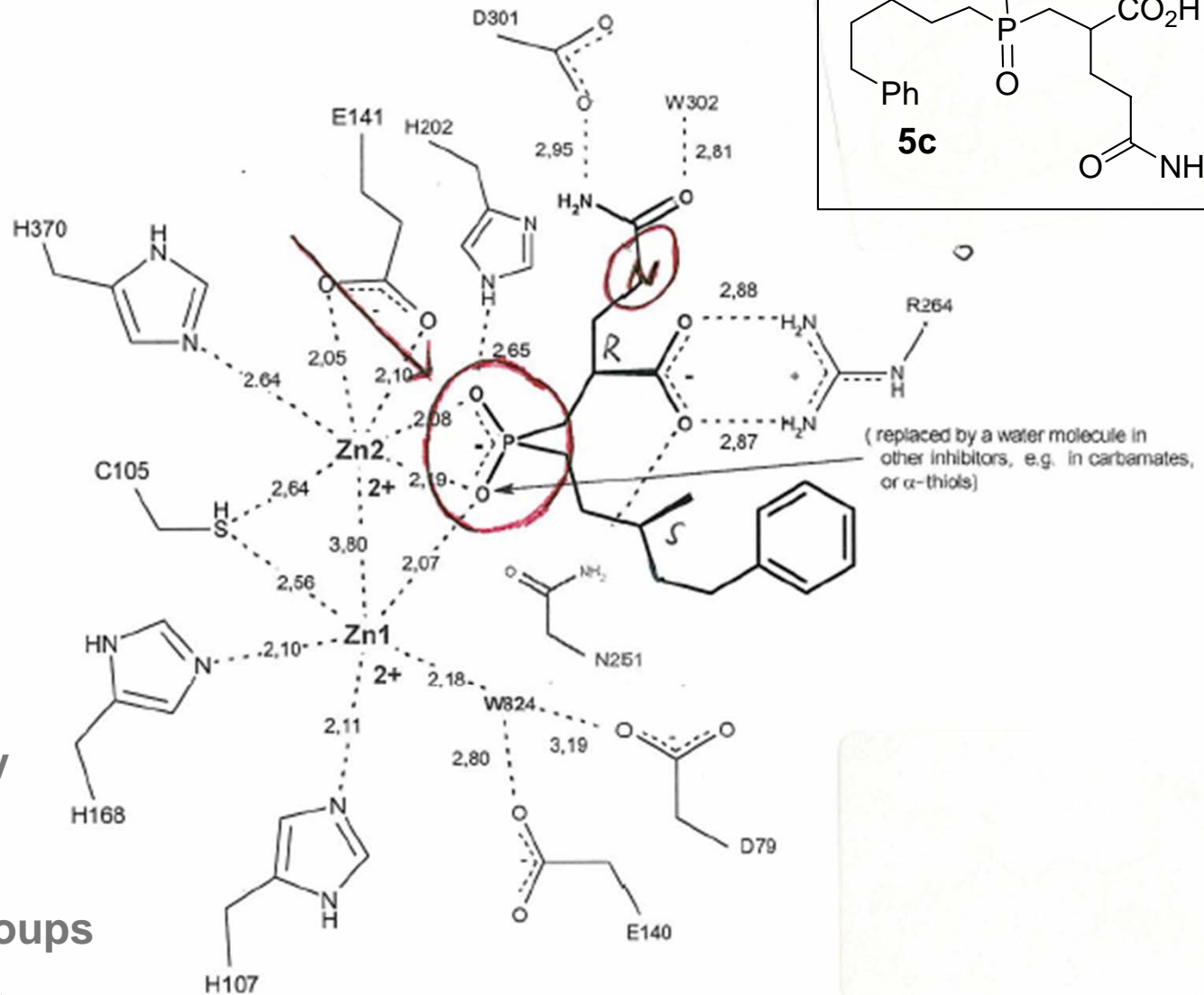
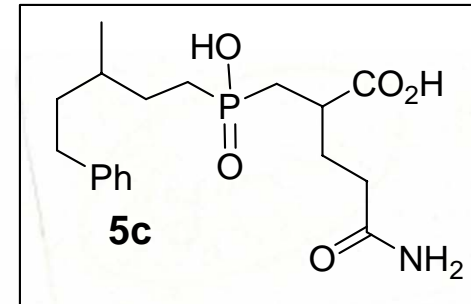
Cristall structure of AMRE with bound inhibitor

- The recombinant enzyme was co-cristallised with the most potent enzyme inhibitor
- Cristal structure resolved to 1.75 Å
- Enzyme tetramer with two zinc atoms in each active site



AMRE / Inhibitor Complex: LB / HBs and Zn-interactions at the active Site

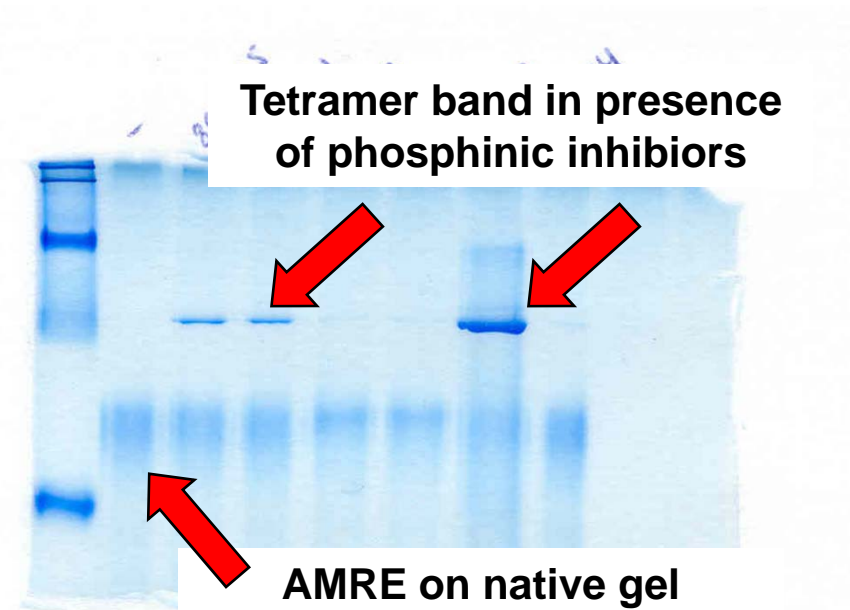
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- Absolute stereochemistry
- Replacement of functional groups

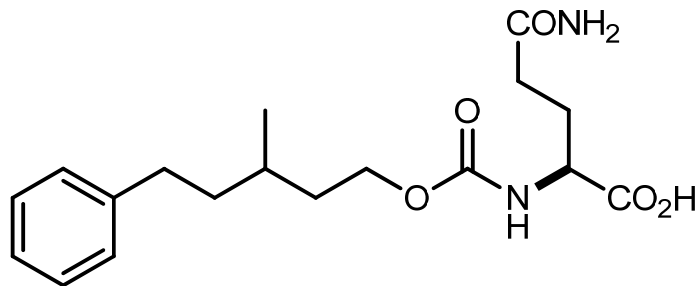
Unusual binding mode of phosphinic inhibitors

- Phosphinic inhibitors are thought to be classical transition state analogues
 - Typically, these are **competitive inhibitors**
 - IC50 is dependent on substrate concentrations (Increasing substrate concentration in test raises IC50)
-
- **This is not the case for the phosphinic inhibitors of AMRE!**
 - Inhibitors give kinetic plots as suicide inhibitors
 - **The phosphinic inhibitors catalyze stable tetramerizations of the enzyme**

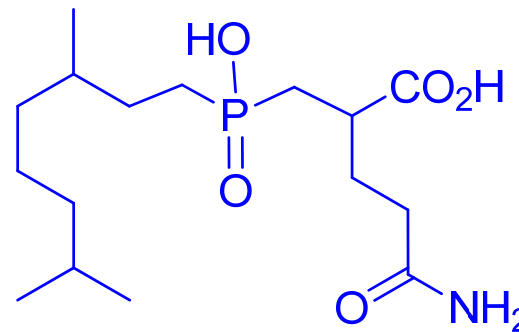


...and the ,clinical' reality

- The ,clinical test' for deodorant ingredients – Sniff test with human volunteers.
- Sequential studies in the same panel with phosphinic lead inhibitor and alternative substrate



GR-85-4027, best alternative substrate

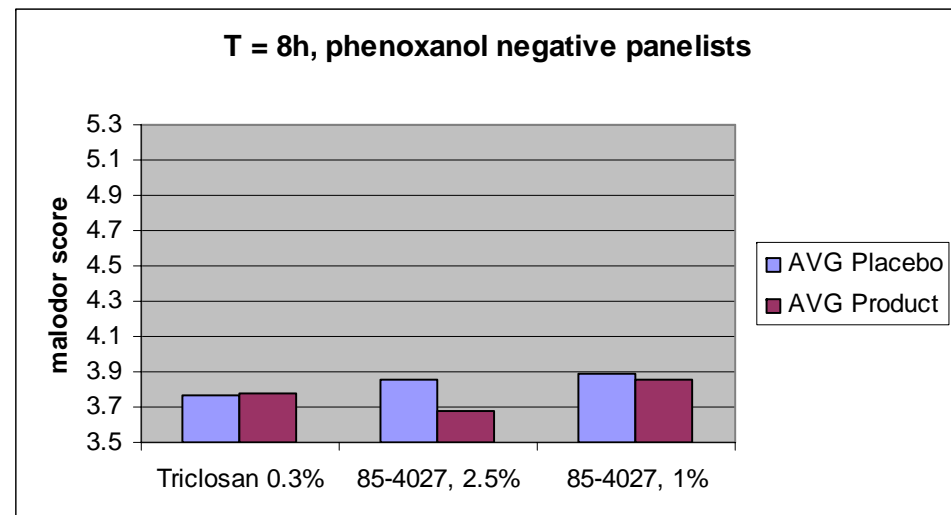
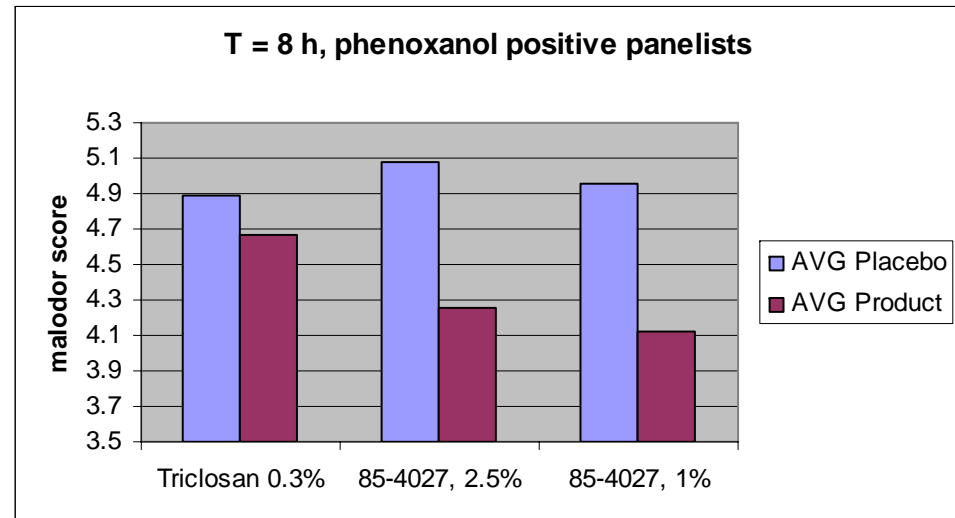


GR-86-3021, best phosphinic inhibitor

Effect of triclosan and alternative substrate 85-4027

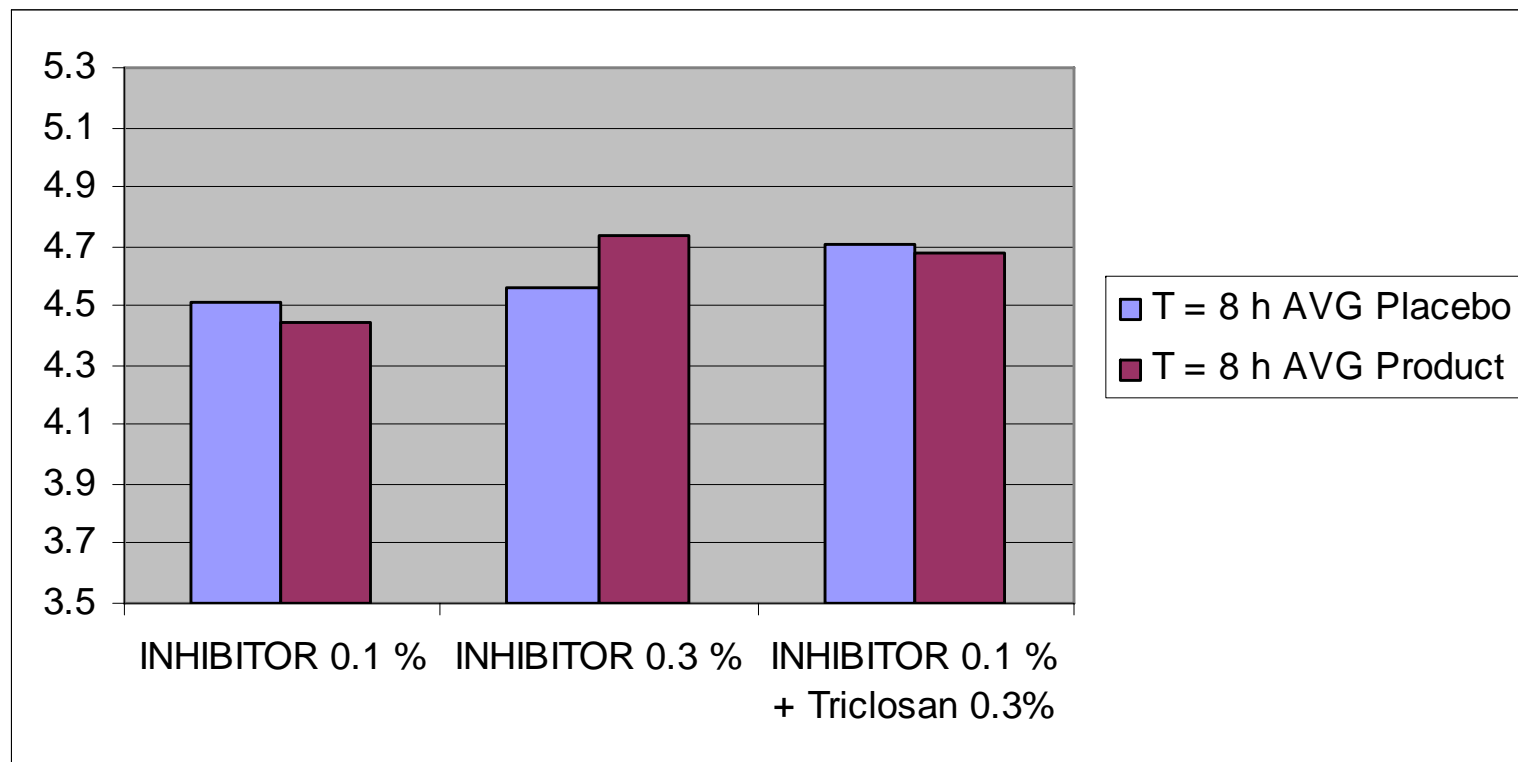
- The effect of 85-4027 is significant in the phenoxanol-positive-panelists with higher malodor score
- The effect on the negative panelists is non-significant

⇒ The product works on those panelists with a real malodor problem



Effect of the phosphinic inhibitor

- NO significant effect of the inhibitor after 8 h in three consecutive studies despite an *in vitro* efficacy of 19 nM IC50!



The ,clinical' reality

- The simple alternative substrate gives significant deodorant benefits
- **Works best on ,high malodor individuals'**
- **Gives a convincing target validation**
- ... But: The most efficient inhibitor failed in the *in vivo* studies
- Reasons?
 - ◆ Poor bioavailability in the axilla?
 - ◆ Specific mode of competitive binding (tetramerization of the enzyme?)

Chemistry and biochemistry of axilla odors - A multidisciplinary project

- **Chemical synthesis** of odorants, precursors, inhibitors and alternative substrates: F. Flachsmann, S. Derrer, T. Granier, S. Elmer, O. Wäckerlig, M. Fournie-Zaluski (Université de Paris, PharmaLeads)
- **GC-MS analysis, NMR:** J. Schmid, G. Brunner
- **Protein purification, enzymatic tests and heterologous expression:** A. Natsch, R. Emter, M. Wasescha, W. Stauch
- **Precursor isolation and LC-MS analytics:** H. Gfeller and G. Acuna
- **Protein cristallisation and structure determination:** A. Douangamath, J. Baker (EvoTec)
- **Threshold determinations and sensory data:** H. Koch
- **Molecular Modelling Studies**
A. Borosy, H.-P. Weber

AND NOW – for something completely different

Why Chinese people do not have this problem
- or

The effect of a SNP in the ABCC11 gene

- SNP (single nucleotide polymorphism) in ABCC11 gene:
White earwax instead of yellow earwax.
- White earwax is known to correlate to low /absent axilla odors
- **This mutation is present in > 95% of the people in central chinese populations and > 70% frequency in larger Asian populations**

Frequency of the ABCC11 negative haplotype

- Black is the functional allele
- In white frequency of the ABCC11 mutation



High frequency of the haplotype with the mutated allele

Worldwide frequency of allele A (open portion in each circle).

Does ABCC11 influence secretion of malodor precursors?

- 25 panelists, genotyped for the SNP based on mouth swab DNA
- All panelists did donate sweat (physical exercise)
- Analysis for sweat precursors (amino-acid conjugates)

- 11 of the panelists have the mutation on both chromosomes
 - ◆ AA
- 7 panelists have one mutated gene, one gene still works
 - ◆ AG
- 7 panelists have no mutation
 - ◆ GG

Genotyp ABCC11	Ethnic population	Secreted amino-acid conjugates (µMol / 2 pads)		
		HMHA-Gln	3M2H-Gln	86-8434
AA	Philippino	n.d. ¹⁾	n.d.	n.d.
AA	Chinese	n.d.	n.d.	n.d.
AA	Philippino	n.d.	n.d.	n.d.
AA	Korean	n.d.	n.d.	n.d.
AA	Chinese	n.d.	n.d.	n.d.
AA	Philippino	n.d.	n.d.	n.d.
AA	Philippino	n.d.	n.d.	n.d.
AA	Philippino	n.d.	n.d.	n.d.
AA	Honkong chinese	n.d.	n.d.	n.d.
AA	Philippino	n.d.	n.d.	n.d.
AA	Philippino	n.d.	n.d.	n.d.
AG	Philippino	1.23	0.17	detectable
AG	Philippino	1.58	0.23	0.041
AG	Philippino	0.06	detectable,	n.d.
AG	Philippino	2.71	0.40	detectable,
AG	Thai	0.89	0.14	detectable,
AG	German	1.18	0.18	0.045
AG	German	0.54	0.10	detectable,
GG	Philippino	0.77	0.13	detectable,
GG	Philippino	0.75	0.11	detectable,
GG	German	1.30	0.19	0.041
GG	German	1.12	0.16	0.038
GG	German	2.65	0.43	0.051
GG	German	0.34	0.09	detectable,
GG	Swiss	0.85	0.18	n.d.

No odor precursor in people with mutation on both chromosomes

One intact gene is sufficient to secrete malodor precursor

ABCC11 – scientific conclusions

- Complete loss of malodor precursor secretion in panelists with two mutant genes
- Second ‚proof of principle‘ or **target validation** – Secretion of the identified malodor substrates correlates to body odors
- High frequency of this evolutionary young mutation: **Positive selection pressure!**
- **Advantage in partner selection** for **low odorant individuals** in ancient asian cultures with long culture of personal hygiene?